Operant behavior to obtain palatable food modifies neuronal plasticity in the brain reward circuit

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Abstract
Palatability enhances food intake by hedonic mechanisms that prevail over caloric necessities. Different studies have demonstrated the role of endogenous cannabinoids in the mesocorticolimbic system in controlling food hedonic value and consumption. We hypothesize that the endogenous cannabinoid system could also be involved in the development of food-induced behavioral alterations, such as food-seeking and binge-eating, by a mechanism that requires neuroplastic changes in the brain reward pathway. For this purpose, we evaluated the role of the CB\textsubscript{1} cannabinoid receptor (CB\textsubscript{1}-R) in the behavioral and neuroplastic changes induced by operant training for standard, highly caloric or highly palatable isocaloric food using different genetics, viral and pharmacological approaches. Neuroplasticity was evaluated by measuring changes in dendritic spine density in neurons previously labeled with the dye DiI. Only operant training to obtain highly palatable isocaloric food induced neuroplastic changes in neurons of the nucleus accumbens shell and prefrontal cortex that were associated to changes in food-seeking behavior. These behavioral and neuroplastic modifications induced by highly palatable isocaloric food were dependent on the activity of the CB\textsubscript{1}-R. Neuroplastic changes induced by highly palatable isocaloric food are similar to those produced by some drugs of abuse and may be crucial in the alteration of food-seeking behavior leading to overweight and obesity.

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1. Introduction

The availability of highly palatable food is a crucial factor that promotes overeating in developed countries (Saper et al., 2002) and finally can lead to overweight, obesity and associated illnesses. The prevalence of obesity has increased substantially since the mid-twentieth century in virtually every country (Caballero, 2007), and obesity prevalence in adults has doubled in the US since 1980, where 34% of US adults are obese (Khan et al., 2009).

Overeating behavior shares similarities with the loss of control and compulsive taking behavior observed in drug-addicts. Thus, deficiencies in inhibitory capacity have been reported in obese humans with excessive food intake (Nederkoorn et al., 2006; Rosval et al., 2006) that are similar to the elevated impulsivity leading to relapse in drug addicts (Moeller et al., 2001). In addition, overconsumption of highly caloric food produces neuroadaptive changes in the brain reward system that can drive the development of compulsive eating (Johnson and Kenny, 2010). Food, like drugs of abuse, activates the mesocorticolimbic system, which mediates the hedonic and motivational aspects of different rewarding stimuli. Repeated activation of this brain pathway may lead to neuroadaptive changes and structural reorganization that could participate in the behavioral alterations promoted by drugs of abuse (Russo et al., 2010). Early studies showed a correlation between changes in neuroplasticity and locomotor sensitization induced by cocaine (Li et al., 2004), although more recent studies have reported conflicting results (for review see Russo et al., 2010). It is currently unclear whether similar neuroplastic alterations occur with repeated exposure to natural rewards. Indeed, most studies have failed to observe changes in structural plasticity in the mesocorticolimbic system induced by food (Robinson et al., 2001; Crombag et al., 2005), although another natural reward, sex, has been recently reported to modify dendritic morphology in the reward circuit (Pitchers et al., 2010).

The endogenous cannabinoid system is a key modulator of the hedonic value of natural rewards (Di Marzo and Matias, 2005) and drugs of abuse (Maldonado et al., 2006). These rewarding responses seem to be mediated, at least in part, by the activation of CB1-R in the mesocorticolimbic system (Maldonado et al., 2006). CB1-R are also involved in mediating neuroplastic changes induced by drugs of abuse (Ballesteros-Yáñez et al., 2007). Considering the key role of CB1-R in food rewarding effects, these receptors could also be involved in any possible neuroplasticity change induced by the exposure to a natural reward.

In this study, we have investigated the ability of palatable food-induced seeking behavior to promote neuroplasticity and the possible involvement of CB1-R in these changes. For this purpose, CB1 knockout mice (CB1−/−) and wild-type littermates (CB1+/+) were trained to lever-press to obtain standard, highly caloric or highly palatable isocaloric pellets and the changes in morphological plasticity were evaluated. We found that operant behavior to obtain highly palatable food produced changes in structural plasticity in specific areas of the mesocorticolimbic system in CB1+/+, but not in CB1−/−.

2. Experimental procedures

2.1. Animals

The experiments were carried out in male CB1−/− and CB1+/+ littermates from 8 to 12 weeks old at the beginning of the experiments. The generation of CB1−/− and CB1+/+ was described previously (Zimmer et al., 1999). Briefly, homozygous CB1−/− and CB1+/− were bred by back-crossing of chimeric and heterozygous animals to C57BL6/J and interbreeding of heterozygous animals for at least 10 generations in order to obtain a pure C57BL6/J background. The animals were individually housed and maintained in a controlled temperature (21 ± 1 °C) and humidity (55 ± 10%) room with a 12:12-h reversed light/dark cycle (on at 8 p.m. and off at 8 a.m.). All the experiments were performed during the dark phase of the dark/light cycle. Animals were habituated to the experimental room and handled for one week before starting the experiments with ad libitum access to standard chow and water. All animal procedures were conducted in accordance with the standard ethical guidelines (European Communities Directive 86/ 60/EEC, Animal Welfare Assurance #A5388-01, IACUC Approval Date 06/08/2009) and approved by the local ethical committee (Comité Étique d’Experimentation Animal—Institut Municipal d’Assistance Sanitaire—Université Pompeu Fabra).

2.2. Acquisition of operant responding maintained by food

Operant responding maintained by food was evaluated in mouse operant chambers (Model ENV-307A-CT, Med. Associates, Georgia, VT, USA). The chambers were made of aluminum and acrylic, had grid floors (EVV-414, Med. Associates Inc., St. Albans, USA), and were housed in sound and light-attenuated boxes equipped with fans to provide ventilation and white noise. The chambers were equipped with two retractable levers, one randomly selected as the active and the other as the inactive. Pressing on the active lever resulted in a pellet delivery (standard, highly caloric or highly palatable isocaloric pellic) together with a stimulus-light during 2 s (associated-cue), while pressing on the inactive lever had no consequences. A food dispenser equidistant between the two levers permitted delivery of food pellets when required. The beginning of the each operant responding session was signaled by turning on a house light placed on the ceiling of the box for 3 s that was then turned off during the remaining duration of the session. The side of the active and inactive lever was counterbalanced between animals. Each session started with a priming delivery of one pellet. A time-out period of 10 s was established after each pellet delivery. During this period, the cue-light was off and no reinforcer was provided after responding on the active lever. Responses on the active lever and all the responses performed during the time-out period were also recorded. The session was terminated after 100 reinforcers were delivered or after 1 h, whichever occurred first. One hour daily sessions were conducted seven days per week during a period of 41 days. The animals were food deprived five days before starting sessions to maintain their weight at 90% of their ad libitum initial weight adjusted for growth, and this food restriction regime was maintained during the first 10 sessions of the operant behavior training to permit the appropriate acquisition of the task. Additional standard and highly palatable isocaloric-yoked groups were included. These groups were subjected to the same experimental conditions except that no reinforcer and no cue-light were presented after pressing in any of the two levers exposed in the operant chamber. However, standard and highly palatable isocaloric-yoked groups received passively the same amount of pellets and with the same frequency as the CB1+/+ that were trained to obtain them.

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The training sessions started with a fixed-ratio 1 (FR1) schedule of reinforcement during 15 days (during the first 10 days mice were food deprived) where one active lever-press resulted in one food pellet. A minimum of 12 CB1+/+ on each experimental group (included highly palatable isocaloric-yoked animals) were sacrificed after the exposure to the 10th session, to evaluate the effects of short term exposure to different types of food on structural plasticity and molecular changes. The rest of the animals were then fed ad libitum during the remaining operant behavior training. Operant training on FR1 was followed by a period of 10 days with a fixed ratio 5 (FR5) schedule (five active lever-presses were required to obtain one pellet). Mice were then exposed to a progressive ratio (PR) schedule in which the response requirement to earn one pellet escalated according to the following series: 1—2—3—5—12—18—27—40—60—90—135—200—300—450—675—1000. The PR session lasted for 4 h or until mice stopped responding for at least 1 h, and was performed only once. The breaking point was determined in each animal as the last response ratio completed. Finally, mice were exposed to a FR5 schedule for 15 additional days. After each session, mice were returned to their home-cages. The chambers were cleaned at the end of each session to remove the presence of odor of the previous mouse. Therefore, the schedule of the experiment is summarized as follows:

<table>
<thead>
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<th>Deprivation</th>
<th>Ad libitum</th>
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<td>FR1</td>
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<td>15 days</td>
<td>FR5</td>
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As previously described (Barbano et al., 2009), the criteria for the achievement of the operant responding were acquired when all of the following conditions were met: (i) mice maintained a stable responding with less than 20% deviation from the mean of the total number of reinforcers earned in three consecutive sessions (80% of stability); (ii) at least 75% responding on the active hole, and (iii) a minimum of 10 reinforcers per session. All the mice included in this study have achieved these criteria in all the experimental phases.

2.3. Food pellets

During the operant experimental sessions, animals were presented with 20 mg dustless precision standard pellets (TestDiet, Richmond, IN, USA), highly palatable isocaloric pellets (TestDiet, Richmond, IN, USA) or highly palatable pellets (Bio-serv, Frenchtown, NJ, USA). The standard pellet formula was similar to the standard maintenance diet provided to mice in their home cage (24.1% protein, 10.4% fat, 65.5% carbohydrate, with a caloric value of 3.30 kcal/g). Highly palatable isocaloric pellets presented similar caloric value to standard pellets (20.5% protein, 12.7% fat, 66.8% carbohydrate, with a caloric value of 3.48 kcal/g) with some differences in their composition. Thus, highly palatable isocaloric pellets were modified by the addition of chocolate flavor (2% pure unsweetened cocoa), and although the carbohydrate content was similar in standard (65.5%) and highly palatable isocaloric pellets (66.8%), the proportion of sugars within this carbohydrate content was different: sucrose content in highly palatable isocaloric pellets was 50.11% of the total carbohydrates, whereas in standard pellets it was only 3.09%. This different composition together with the cocoa content made highly palatable isocaloric pellets much more palatable than standard pellets. Finally, we also used a highly caloric fat-enriched formula (14% protein, 60% fat, 26% carbohydrate, with a caloric value of 5.32 kcal/g). These pellets were presented only during the operant behavior sessions. Otherwise, animals were maintained on standard chow for their daily food intake.

2.4. Ballistic labeling with the fluorescent dye DiI

See supplementary experimental procedures.

2.5. Dendritic spine analysis

Individual medium spiny neurons in the NAc and pyramidal neurons from the mPFC were chosen for spine analysis based on several criteria, as described previously (Lee et al., 2006): (i) there was minimal or no overlap with other labeled cells to ensure that processes from different cells would not be confused, (ii) at least three primary dendrites needed to be visible for cells to be used for analysis and (iii) distal dendrites (from secondary dendrites to terminal dendrites) were examined. Dendrites from medium spiny neurons in the core and shell of the NAC (from bregma 1.54 to bregma 1.10) and basilar dendrites of pyramidal neurons taken predominantly from the prelimbic and infralimbic areas of the mPFC (from bregma 1.98 to bregma 1.70) were analyzed.

To calculate spine density, a length of dendrite (at least 20 μm long) was traced by using a confocal microscope (Zeiss LSM 510, Germany) with an oil immersion lens (40 ×). All images of dendrites were taken at different z levels (0.3 μm depth intervals) to examine the morphology of dendritic spines. All measurements were made using IMAGE J analysis software. Protrusions from dendrites were classified into five types based on their morphology: class 1 protrusions, also called stubby protruberances were between 0.5 and 1.25 μm in length and lacked a large spine head, and did not appear to have a neck; class 2, or mushroom-shaped spines were between 0.5 and 1.25 μm in length and were characterized by a short neck and large spine head; class 3, or thin spines ranged between 1.25 and 3.0 μm and had elongated spine necks with small heads; class 4, or wide spine were between 0.5 and 1.25 μm in length and were characterized by a large neck and a large spine head; and class 5 or branched spine ranged between 1.25 and 3.0 μm and had elongated spine necks with two or more spine heads. Quantification of dendritic spine densities was performed in blind conditions.

2.6. Acute bilateral intraNAc microinjection of rimonabant

See supplementary experimental procedures.

2.7. Design and construction of AAV vectors and viral production and purification

See supplementary experimental procedures.

2.8. Bilateral intraNAc administration of AAV-scrambled or AAV-sh CB1

See supplementary experimental procedures.

2.9. Tissue preparation and immunofluorescence analysis

See supplementary experimental procedures.

2.10. Immunoblot analysis

See supplementary experimental procedures.

2.11. Statistical analysis

The number of food pellets consumed and the active responses during the time-out period were analyzed by repeated measures two-way ANOVA with genotype (between subjects) and day (within subjects) as factors of variation, followed by corresponding post-hoc analysis when required (Dunnett’s test).
Immunoblot data were analyzed by one-way ANOVA between subjects. Structural plasticity data were analyzed by a two-way ANOVA with genotype and kind of food as between subject factors of variation, followed by corresponding post-hoc analysis when required (Dunnett’s test). An additional one-way ANOVA between subjects was used to compare yoked CB1+/+ receiving highly palatable isocaloric pellets non-contingently with other groups of CB1+/+ followed by the corresponding post-hoc Dunnett’s analysis when required. All data were analyzed with SPSS software and are expressed as mean ± SEM. Differences were considered significant at p<0.05.

3. Results

3.1. Operant behavior to obtain highly palatable isocaloric food modifies seeking behavior through a CB1-R dependent mechanism

Caloric content and palatability are two important properties of food involved in compulsive food intake (Johnson and Kenny, 2010; Davis and Carter, 2009). The endogenous cannabinoid system, through the activation of CB1-R, plays a key role in modulating food hedonic value. Therefore, we evaluated the behavioral consequences of prolonged training to obtain palatable food and the possible involvement of CB1-R in these responses. CB1−/− and CB1+/+ were trained in an operant paradigm to obtain different kinds of food pellets: control standard diet (3.30 kcal/g), highly caloric food rich in fat (high-fat pellets, 5.32 kcal/g) or an isocaloric highly palatable food (highly palatable isocaloric pellets, 3.48 kcal/g). In fasted conditions, CB1−/− showed a decrease in the consumption and operant responding for standard, highly caloric and highly palatable isocaloric pellets compared with CB1+/+ (Fig. 1A-C). The role of the endocannabinoid system regulating food intake and energy balance in central and peripheral tissues (Cota et al., 2003) could explain these differences (see discussion section).

In fed conditions, no differences in the operant behavior to obtain standard pellets were observed between genotypes (Fig. 1A). However, a decrease in the number of lever-presses to obtain highly palatable isocaloric pellets was observed during the training sessions in fed conditions in CB1−/− when compared to CB1+/+ (Fig. 1C). A progressive ratio (PR) schedule was used to evaluate the motivation of mice for each type of food. Motivation of CB1+/+ to obtain this highly palatable isocaloric food was significantly higher than for standard or highly caloric pellets (Fig. 1D). CB1−/− trained to obtain highly caloric pellets showed an enhancement in the number of lever-presses compared with CB1+/+ (Fig. 1B). In agreement, CB1−/− also showed an increased response for this type of food in the progressive ratio paradigm when compared to CB1+/+ (Fig. 1D). This last difference might be due to the caloric deficit produced by the deletion of the CB1-R since these animals present a lower amount of fat content (Cota et al., 2003) (see discussion section).

The number of lever-presses during the time-out periods that had no rewarding consequences was also evaluated as an indirect measure of food-seeking behavior that can be related to enhanced impulsivity. Training for highly palatable isocaloric pellets, but not for other types of food, increased the number of lever-presses during the time-out period at the beginning of the training period on fixed ratio (FR) 5 in CB1+/+. This response was further enhanced the last day of training on FR5 when compared with the initial training (Fig. 2), suggesting that palatable food progressively enhances this impulsive-like behavior developed during the training period. In contrast, this behavioral consequence of highly palatable isocaloric food training was not observed in CB1−/−, which showed similar responses during the time-out periods when trained for the different kinds of food (Fig. 2).

We also evaluated the time course changes in body weight in both genotypes during the entire experiment. The lean phenotype previously reported in CB1−/− (Cota et al., 2003) was mainly maintained in this study in CB1−/− trained to obtain standard food, although weight differences between genotypes were attenuated in the groups trained to obtain highly caloric or highly palatable isocaloric pellets (data not shown). These results rule out a possible influence of differences in body weight in the changes in lever-pressing and food consumption observed between genotypes.

These results demonstrate that the high palatability of food, but not its caloric content, is associated with an enhanced motivation of mice to obtain food that progressively leads to increased seeking behavior, which can be related to enhanced impulsivity. These responses were absent in CB1−/− showing that the endocannabinoid system plays a critical role in this behavioral alteration.

3.2. Operant behavior to obtain highly palatable isocaloric food modifies structural plasticity in the brain reward circuit

Long-lasting behavioral changes induced by drugs of abuse have been related to persistent changes in dendritic spines density in key areas of the brain reward system, such as the NAc core, NAc shell and the mPFC (Russo et al., 2010). We therefore evaluated whether operant behavioral training to obtain standard, highly caloric or highly palatable isocaloric pellets could also induce structural neuroplastic changes in these brain structures. Immediately after the last training session, animals were sacrificed and their brains processed for ballistic delivery to label whole neurons with the dye Dil. We did not observe any modification in dendritic spine density in the NAc core, shell or mPFC of CB1+/+ and CB1−/− trained on the operant lever-pressing to obtain standard food (Fig. 3A and B). In contrast, operant training to obtain highly palatable isocaloric pellets increased dendritic spine density in CB1+/+ in the NAc shell and in a lesser extent in the mPFC (Fig. 3A and B). Interestingly, these structural changes were not observed in the CB1+/+ highly palatable isocaloric-yoked group (Fig. 3D) that consumed similar amount of highly palatable isocaloric pellets than their CB1+/+ master-paired mice (Fig. 3C), revealing that this response was selectively due to the operant training to obtain highly palatable isocaloric food, and not to the passive exposure to this kind of pellets. Moreover, no differences in spine density in any of the brain areas investigated were observed between CB1+/+ trained to lever-press for standard pellets and their yoked-standard counterparts (Fig. 3D) in spite of the different operant performance of these two groups (Fig. 3C). These data

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demonstrate that the operant training by self and the associated motor responses were not correlated to the modifications in spine density observed in CB1 +/+ mice. Additionally, these plasticity changes were absent in mice exposed to a short training period (10 days), demonstrating that they are selectively produced by repeated food operant training (Fig. 3E).

Interestingly, a significant correlation was observed in CB1 +/+ in the last training session between the changes in dendritic spine densities in the NAc shell and mPFC and the number of pellets consumed as well as the number of lever presses during the time-out period (Fig. 4). Unlike the NAc shell and the mPFC, no modifications in dendritic spine density were observed in the NAc core (Fig. 3B), demonstrating differential adaptive changes produced by operant behavior to obtain highly palatable isocaloric food in both subregions of the NAc. Therefore, training to obtain highly palatable isocaloric food produced morphological changes in restricted areas of the brain reward circuit that are correlated to the enhanced seeking behavior produced by this operant training in CB1 +/+ mice.

Dendritic spines are classified into different groups depending on their morphology (stubby, thin, branched, wide and mushroom-type spines) and can undergo remodeling, that modifies their functionality (Bourne and Harris, 2007). Therefore, we also investigated the types of dendritic spines that were modified by operant behavior to obtain highly palatable isocaloric food in both subregions of the NAc. Therefore, training to obtain highly palatable isocaloric food produced morphological changes in restricted areas of the brain reward circuit that are correlated to the enhanced seeking behavior produced by this operant training in CB1 +/+ mice.

**Fig. 1** Highly palatable isocaloric food strongly promotes operant behavior and enhances motivation for food through a CB1-R dependent mechanism. (A) Daily consumption of standard pellets in CB1 +/+ (black triangle; n=36) and CB1 −/− (white triangle; n=14) mice. (B) Daily consumption of highly caloric pellets in CB1 +/+ (black rectangle; n=29) and CB1 −/− (white rectangle; n=13). (C) Daily consumption of highly palatable isocaloric pellets in CB1 +/+ (black circle; n=37) and CB1 −/− (white circle; n=17). (D) Breaking point reached by CB1 +/+ (black bars) and CB1 −/− (white bars) for each kind of food in a progressive ratio paradigm, as a measure of motivation for food. Data are expressed as mean±s.e.m. ★ ★ ★ ★ p<0.05, ★ ★ p<0.01, ★ ★ ★ p<0.001 (CB1 +/+ vs. CB1 −/−); ☆ ☆ ☆ p<0.001 (comparison vs. standard food).
isocaloric food, but not other kinds of food (Fig. 5). These changes were absent in CB1−/− trained to obtain highly palatable isocaloric food. CB1−/− only showed an enhanced density of branched spines in the NAc when trained with highly caloric or highly palatable isocaloric pellets (Fig. 5). In addition, we also observed a small increase in the density of branched spines in the NAc shell and mPFC as well as in mushroom spines in the NAc shell in CB1+/+ trained with highly palatable isocaloric pellets (Fig. 5 and Table 1). These results demonstrate that operant training to obtain highly palatable isocaloric food produces structural neuroplastic changes due to the modification of specific types of dendritic spines, mainly thin spines, in the brain reward circuit. These structural changes selectively occur in mice showing enhanced seeking behavior after operant training.

3.3. CB1-R in the nucleus accumbens mediates the behavioral and structural changes promoted by highly palatable isocaloric food

Our previous results suggest that CB1-R activity in the mesocorticoolimbic system might regulate highly palatable isocaloric

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food-induced behavioral and neuroplastic changes. We therefore evaluated the effects produced on these changes by the local inhibition of CB1-R in the NAc, the brain area where highly palatable isocaloric food produced the strongest neuroplastic alterations. For this purpose, we first used an AAV9-sh CB1 approach to knock-down the expression of CB1-R in the NAc of CB1+/+. The bilateral injection of AAV9-sh CB1 produced an inhibition of 50% of the total CB1-R in this brain area in CB1+/+ (Fig. 6B). Under these experimental conditions, the inhibition produced by AAV9-sh CB1 was specific of neurons and did not occur in glial cells (Fig. 6C and D). Control mice receiving AAV9-scrambled into the NAc showed the same consumption of highly palatable isocaloric pellets during FR1 and FR5 training, as well as similar responding in the PR paradigm and the time-out periods (Fig. 6E–G) than CB1+/+ in the previous experiment (Fig. 1C and D and Fig. 2). AAV9-sh CB1 treated mice showed a decrease in the consumption of highly palatable isocaloric food (Fig. 6E), the breaking point response in the PR paradigm (Fig. 6F) and the changes promoted by highly palatable isocaloric food on the lever-presses during the time-out periods (Fig. 6G) in comparison with AAV9-scrambled control animals.

We also evaluated the effects of an acute administration of the selective CB1-R antagonist rimonabant locally into the NAc of CB1+/+. In agreement with the previous results, rimonabant bilaterally microinjected into the NAc at the lowest dose (1 μg/site) selectively decreased highly palatable isocaloric pellet consumption (Fig. 7A). Higher doses of rimonabant produced non-specific effects that were not revealed when using the genetic approach in the previous experiments (see Fig. 1), and also decreased the consumption of other kinds of pellets (Fig. 7A). Taken together the results of both experimental approaches, we clearly

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Fig. 3 Neuronal morphological changes induced by highly palatable isocaloric food are mediated by a CB1-R mechanism. (A) Illustration of DiI-labeled dendrites of medium spiny neurons in the NAc shell of CB1+/+ and CB1−/− trained to obtain standard, highly caloric or highly palatable isocaloric pellets. (B) Total dendritic spines density in neurons from the NAc shell, core and mPFC of CB1+/+ (black bars) and CB1−/− (white bars) after repeated operant training to obtain standard, highly caloric or highly palatable isocaloric pellets. Data represents the average of 7-8 mice per experiment group and a total of 6-12 dendrites per animal and brain area. No more than 2 dendrites were evaluated from the same neuron. (C) Number of pellets consumed and number of active lever pressings per hour in CB1+/+ trained to obtain standard or highly palatable isocaloric pellets and their corresponding yoked-control groups (n=20-28 animals per group). (D) Total dendritic spines density in neurons from the NAc shell, core and mPFC of CB1+/+ trained to obtain standard or highly palatable isocaloric pellets and their corresponding yoked-control groups (n=7-9 animals per group). A total of 6-12 dendrites per animal and brain area were evaluated. No more than 2 dendrites from the same neuron were included. (E) Total dendritic spine density in CB1+/+ exposed to operant training for standard, highly caloric or highly palatable isocaloric food for a short period of 10 days (n=6 mice/group). A total of 6-12 dendrites per animal and brain area were evaluated. No more than 2 dendrites from the same neuron were included. Data are expressed as mean±s.e.m. ★ p<0.05 (CB1+/+ vs. CB1−/−); ★★ p<0.01, ★★★ p<0.001 (comparison between contingent and yoked control group).
demonstrate the selective involvement of CB1-R in the NAc in the behavioral responses induced by highly palatable isocaloric food. These behavioral results obtained after down-regulation of CB1-R in the NAc were similar to those obtained in CB1−/−, ruling out possible compensatory mechanism in the knockout mice that could interfere with these responses.

Down-regulation of CB1-R in the NAc by AAV9-sh CB1 also attenuated the changes promoted by highly palatable isocaloric food in dendritic spine density in the NAc shell that were revealed in control AAV9-scrambled mice. Indeed, the dendritic spine morphology in these control mice (Fig. 6H-J) was similar than in CB1+/+ in the previous experiment (Fig. 3). This attenuation was mainly due to a decrease in the density of thin, branched and mushroom subtypes of spines (Fig. 6J). These results and the similarities to those previously revealed in CB1−/− demonstrate the specific involvement of CB1-R in the NAc in food-induced neuroplastic changes, and the cause-relationship between behavioral and morphological changes induced by highly palatable isocaloric food.

4. Discussion

In this study, we revealed for the first time that the hedonic value of food, but not its caloric content, modifies structural plasticity in restricted areas of the corticolimbic system. These morphological changes selectively occur in mice showing enhanced food-seeking responses and associated impulsivity-like behavior. Using knock-out, pharmacological and viral approaches, we also demonstrated that CB1-R are necessary to produce these behavioral and morphological changes induced by reinforced learning with highly palatable isocaloric food.

In agreement with previous studies (see for review Saper et al., 2002), we observed that food palatability strongly
promotes operant behavior and represents the key factor for food motivation. Moreover, mice working for highly palatable isocaloric pellets progressively increased their operant responses even when no reward can be obtained, suggesting an enhancement of food-seeking behavior that can be related to an enhanced impulsivity. Accordingly, escalation of palatable food seeking behavior has been correlated with elevated impulsivity (Diergaarde et al., 2009), and intermittent access to this kind of food leads to behavioral and neurochemical changes that resemble those produced by drugs of abuse (Avena et al., 2008; Belin et al., 2008). Therefore, chronic exposure to highly palatable isocaloric food enhances reward-related seeking responses and impulsive-like behavior, which could account for progressive food overconsumption and the development of food-seeking and binge-eating. Our operant paradigm is an useful model to reveal these behavioral alterations induced by highly palatable isocaloric food and to study their neurobiological substrate. A behavioral alteration leading to loss of control and addictive-like behavior has also been recently reported in rodents exposed to highly palatable food (Johnson and Kenny, 2010).

Repeated exposure to drugs of abuse modifies structural plasticity in the brain reward system, which may underlie some addictive behaviors (Russo et al., 2010). Previous studies have mainly focused on the morphology of dendritic spines since the majority of excitatory synaptic inputs involved in neuroplastic modifications occur in these structures (Harris and Kater, 1994). We therefore evaluated whether plastic changes similar to those produced by drugs of abuse also occurred in mice exposed to the different kinds of food in our operant paradigm. We found that operant training with highly palatable isocaloric food increased spine density in the NAc shell and in a lesser extent in the mPFC, two brain regions crucial for regulating the hedonic aspects of food (Baldo and Kelley, 2007; Mahler et al., 2007), decision-making and impulsive responses (Volkow and Fowler, 2000). Interestingly, similar structural changes induced by chronic exposure to psychostimulants or nicotine have been hypothesized to participate in the development of addictive behavior, although conflicting results have been recently reported (Russo et al., 2010).

We hypothesize that the increased spine density observed in the NAc shell and mPFC might be responsible for the abnormal behavioral responses observed in mice trained to obtain highly palatable isocaloric pellets, including enhanced seeking-behavior and related impulsivity. Overtraining and the motor responses associated to operant training were not responsible of the changes in dendritic spine densities observed in CB1+/+ since no differences in structural

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plasticity were observed between CB1+/+ trained for standard pellets and their corresponding yoked animals, in spite of the different operant performance of these two groups. Previous studies did not observe changes in structural plasticity 30 days after operant training with highly palatable isocaloric food in rats (Crombag et al., 2005). However, drugs of abuse also extensively modify dendritic spine density in the NAc core (Russo et al., 2010), suggesting that morphological alterations in this region might depend on the hedonic value of food (Pecina and Berridge, 2005). However, drugs of abuse also extensively modify dendritic spine density in the NAc core (Russo et al., 2010), suggesting that morphological alterations in this region might play a more prominent role in mediating the effects of drugs of abuse than palatable food (McFarland et al., 2003).

We have also investigated the possible involvement of CB1-R in the behavioral and morphological effects of highly palatable isocaloric food considering the crucial role of these receptors in food hedonic value (Mahler et al., 2007). The constitutive deletion of CB1-R decreased operant behavior and the motivation to obtain highly palatable isocaloric food, without affecting the operant responses to obtain standard pellets. The behavioral alterations produced by highly palatable isocaloric food in CB1+/+ were also absent in CB1−/−. To evaluate the specific participation of CB1-R in the NAc and to rule out possible compensatory changes in the CB1−/−, we used pharmacological and viral approaches to locally decrease CB1-R activity in the NAc in CB1+/+. The acute pharmacological blockade of CB1-R in the NAc as well as the chronic down-regulation of these receptors by intraNAc microinjection of AAV9-sh CB1 decreased the behavioral

### Table 1  Dendritic spine density of mushroom-, stubby- or wide-type of dendritic spines in CB1+/+ and CB1−/−.

<table>
<thead>
<tr>
<th></th>
<th>Mushroom</th>
<th>Stubby</th>
<th>Wide</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Shell</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CB1+/+</td>
<td>2.61±0.43</td>
<td>1.67±0.60</td>
<td>1.02±0.15</td>
</tr>
<tr>
<td>Fat</td>
<td>2.85±0.53</td>
<td>1.51±0.29</td>
<td>1.24±0.14</td>
</tr>
<tr>
<td>Palatable</td>
<td>2.74±0.37</td>
<td>1.57±0.63</td>
<td>1.21±0.19</td>
</tr>
<tr>
<td>Yoked palatable</td>
<td>1.47±0.31</td>
<td>3.26±0.59</td>
<td>1.14±0.16</td>
</tr>
<tr>
<td>CB1−/−</td>
<td>2.56±0.47</td>
<td>1.98±0.43</td>
<td>1.29±0.22</td>
</tr>
<tr>
<td>Fat</td>
<td>2.87±0.28</td>
<td>1.47±0.25</td>
<td>1.09±0.17</td>
</tr>
<tr>
<td>Palatable</td>
<td>2.72±0.25</td>
<td>1.44±0.14</td>
<td>1.27±0.20</td>
</tr>
<tr>
<td><strong>Core</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CB1+/+</td>
<td>2.78±0.44</td>
<td>0.94±0.15</td>
<td>0.96±0.26</td>
</tr>
<tr>
<td>Fat</td>
<td>2.96±0.39</td>
<td>1.8±0.67</td>
<td>1.05±0.12</td>
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<tr>
<td>Palatable</td>
<td>2.84±0.61</td>
<td>1.71±0.61</td>
<td>1.07±0.16</td>
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<tr>
<td>Yoked palatable</td>
<td>2.2±0.52</td>
<td>3.54±0.75</td>
<td>1.28±0.12</td>
</tr>
<tr>
<td>CB1−/−</td>
<td>2.61±0.36</td>
<td>1.56±0.61</td>
<td>0.94±0.10</td>
</tr>
<tr>
<td>Fat</td>
<td>3.73±0.35</td>
<td>1.47±0.19</td>
<td>0.99±0.18</td>
</tr>
<tr>
<td>Palatable</td>
<td>2.94±0.3</td>
<td>1.09±0.11</td>
<td>1.24±0.13</td>
</tr>
<tr>
<td><strong>mPFC</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CB1+/+</td>
<td>2.18±0.29</td>
<td>2.46±0.59</td>
<td>1.19±0.12</td>
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<tr>
<td>Fat</td>
<td>2.03±0.36</td>
<td>2.02±0.51</td>
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<td>Yoked palatable</td>
<td>2.01±0.39</td>
<td>3.08±0.66</td>
<td>1.17±0.06</td>
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<tr>
<td>CB1−/−</td>
<td>3.19±0.35</td>
<td>1.42±0.16</td>
<td>1.13±0.17</td>
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<tr>
<td>Fat</td>
<td>2.2±0.37</td>
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<td>0.83±0.10</td>
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<tr>
<td>Palatable</td>
<td>3.14±0.48</td>
<td>1.09±0.07</td>
<td>0.98±0.19</td>
</tr>
</tbody>
</table>

*Indicates significant differences between “contingent highly palatable isocaloric” and “highly palatable isocaloric-yoked groups”.

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(pharmacological and viral approaches) and structural plasticity (viral approach) responses induced by highly palatable isocaloric food in a similar manner that was observed in CB1+/−. These results demonstrate the specific involvement of CB1-R in the NAc in these behavioral and morphological changes promoted by highly palatable isocaloric food. Interestingly, other studies have reported that the endogenous cannabinoid system regulates behavioral aspects related to the loss of control over drug intake (Maldonado et al., 2006), and modulates impulsivity in a stop signal task (McDonald et al., 2003). Our results extend these observations and suggest that CB1-R also modulate seeking behavior and related impulsivity promoted by the exposure to highly palatable isocaloric food. On the other hand, CB1+/− showed an enhanced operant response and motivation to obtain highly caloric pellets. This observation agrees with several studies showing that CB1+/− have a lower amount of fat content (Cota et al., 2003) and would require more
caloric intake than CB1+/+. A decreased lipid storage in adipocytes in CB1−/− might be responsible (Cota et al., 2003), at least in part, of the lean phenotype and increased highly caloric pellet consumption observed in these knockout animals. In addition, a recent study has shown that endocannabinoid-induced CB1-R activation in the small intestine contributes to orosensory-mediated induction of fat intake (DiPatrizio et al., 2011). Our results and those recent findings support the role of CB1-R in the regulation of highly caloric food intake.

The acquisition of operant responding is a learning/memory process that can lead to the change of the shape and structure of the dendritic spines, which also modifies their functionality (Bourne and Harris, 2007). In agreement, predominant enhancement in the proportion of thin spines in the NAc shell and mPFC was observed in mice trained to obtain highly palatable isocaloric food, but not other kinds of food. These changes were mediated by CB1-R in the NAc since they were absent in CB1−/− and CB1+/+ receiving AAV9-sh CB1 into this brain structure. Thin spines are considered transient and highly plastic structures that can both stabilize and transform into more mature spines in response to behavioral changes and during learning processes by incorporating AMPA receptors (Bourne and Harris, 2007) or can shrink and finally disappear. Hence, the enhancement of thin spines induced by operant training to obtain highly palatable isocaloric food could increase the strength of specific synapses in the NAc shell and mPFC, and promote the behavioral abnormalities observed during this operant training. Although operant training with highly palatable isocaloric food mainly enhanced the density of thin spines, the microinjection of AAV9-sh CB1 into the NAc also decreased the density of other kind of spines. Thus, the generation of stabilized spines appears to represent a plasticity change associated to highly palatable isocaloric food operant learning with the participation of CB1-R.

In conclusion, we reveal behavioral alterations promoted by operant training to obtain highly palatable isocaloric food that are associated to neuroplastic changes. Similar morphological alterations have been reported in these brain structures after exposure to drugs of abuse, which play a crucial role in addictive behavior. Therefore, these neural changes induced by highly palatable isocaloric food could represent a biological substrate to explain some of the alterations in food-seeking behavior that could eventually lead to eating-related disorders and promote obesity. Our results also provide an important advance in the understanding of the common links between food-seeking and drug addiction.

Fig. 7 Acute inhibition of CB1-R activity in the NAc of CB1+/+ decreases operant responding and pellet consumption. (A) Histogram showing the total number of standard, highly caloric and highly palatable isocaloric pellets consumed before treatment after surgery (mean of the last three sessions before treatment), and after bilateral injection into the NAc of vehicle or the different doses of rimonabant. n=6–9 mice per experimental group. (B) Schematic diagrams adapted from the mouse brain atlas of Paxinos and Franklin (2001) showing representative injection sites of rimonabant in the NAc. Black dots indicate locations of cannula tips. Each image indicates the distance from Bregma. Data are expressed as mean±s.e.m. ★★p<0.05, ★★★p<0.01, ★★★★p<0.001 (comparison vs. standard food); #p<0.05; ##p<0.01 (vehicle vs. different doses).

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Role of founding source

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Contributors

T.G. conducted the behavioral and structural plasticity studies and helped in the interpretation and manuscript writing. L.C. conducted the behavioral and structural plasticity studies. E.S. and G.G. conducted immunohistochemical studies. E.A. and F.B. were involved in the generation of the AAV9. G.F. funded part of the project. A.M. supervised part of the project. E.V. conducted immunohistochemical studies, participated in experimental design and helped in the interpretation and manuscript writing. R.M. participated in the experimental design, helped in the interpretation and manuscript writing and funded part of the project. M.M. participated in the experimental design, helped in the interpretation and manuscript writing.

Conflicts of interest

All authors declare no biomedical financial interest or potential conflicts of interests.

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Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at http://dx.doi.org/10.1016/j.euroneuro.2012.04.004.

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